## S2.11 Structure of photosystem I and its natural electron acceptor ferredoxin in co-crystals at 3.8 Å resolution

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Photosystem I is a large membrane protein complex that catalyzes the first step of light reactions in photosynthesis. The molecular structure of this complex is solved to atomic resolution (2.5 A). Ferredoxin (Fd) acts as the natural electron acceptor of Photosystem I and mediates the electron transfer from Photosystem I to the FNR, where finally NADP+ is reduced to NADPH. The aim of our studies is to unravel the interaction between Photosystem I and ferredoxin at atomic detail by co-crystallization of Photosystem I with ferredoxin. The trimer of Photosystem I has a MW of 1 056 kDa compared with 10 kDa for ferredoxin. The phase was solved by a combination of molecular replacement and heavy atom anomalous diffraction. The position of ferredoxin was predicted by modeling the docking of ferredoxin to PS I and confirmed by omit mapping. The space group has been determined to be P21 with a=214.5, b=235.6, c=261.2 A and alpha=90.0, beta=100.47, gamma=90.0. The R-factor of the current model is 21.8% and the Rfree is 34.2%. In the asymmetric unit are six PS I and six Fd. Docking and binding of Fd to PS I and electron transfer are now discussed in detail with respect to the co-crystal structure with the distances to the subunits PsaA and PsaB and the three extrinsic subunits Psa C, D and E to the Fe<sub>4</sub>S<sub>4</sub> and Fe<sub>2</sub>S<sub>2</sub> clusters.

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## S2.12 Growth and photosynthetic performance of the ricefield cyanobacterium *Anabaena cylindrica* to the herbicide bentazon

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Bentazon is a selective herbicide recommended for integrated rice weed management and acts by binding to the exchangeable quinone at the photosystem II (PS II) reaction centre. However, its precise molecular mechanism of inhibition has not been yet well characterized and its phytotoxic effects remain unexplained. In this study, the effects of bentazon on dry weight yield, chlorophyll a content, photosynthesis (complementary analysis of O2 evolution and of quantum efficiency of PS II) and respiration were studied in Anabaena cylindrica, a cyanobacterium isolated from Portuguese rice fields, in a time- and dose-dependent exposure throughout 72 h. Higher bentazon concentrations induced a significant decline on biomass yield with time. Whereas concentrations ranging from 0.75 to 2 mM did not significantly modified chlorophyll a content with time, photosynthesis (O2 evolution) and respiration (O2 consumption) were severely inhibited in a time and dose response manner, particularly with higher concentrations. Bentazon also significantly reduced the fluorescence parameters  $F_v/F_m$ ,  $\Phi_{PSII}$  and qP, as indicators of photosynthetic performance. Since A. cylindrica

is a primary source of aquatic food web and an important biofertilizer for rice cultivation, its protection from potential residual effects of bentazon is essential for enriched local soil fertility.

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#### (S3) Membrane transporters symposium lecture abstracts

#### S3/1 An ancient look at UCP1

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Brown adipose tissue serves as a thermogenic organ in placental mammals to defend body temperature in the cold by nonshivering thermogenesis. The thermogenic function of brown adipose tissue is enabled by several specialised features on the organ as well as on the cellular level, including dense sympathetic innervation and vascularisation, high lipolytic capacity and mitochondrial density and the unique expression of uncoupling protein 1 (UCP1). This mitochondrial carrier protein is inserted into the inner mitochondrial membrane and stimulates maximum mitochondrial respiration by dissipating protonmotive force as heat. Studies in knockout mice have clearly demonstrated that UCP1 is essential for nonshivering thermogenesis in brown adipose tissue. For a long time it had been presumed that brown adipose tissue and UCP1 emerged in placental mammals providing them with a unique advantage to survive in the cold. Our subsequent discoveries of UCP1 orthologues in ectotherm vertebrates and marsupials clearly refute this presumption. We can now initiate comparative studies on the structure-function relationships in UCP1 orthologues from different vertebrates to elucidate when during vertebrate evolution UCP1 gained the biochemical properties required for nonshivering thermogenesis.

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### S3/2 Structural studies on bacterial and mammalian transporters

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Membrane transporters that transduce free energy stored in electrochemical ion gradients into a concentration gradient are a major class of membrane proteins. We have been studying the structure and mechanism of membrane transporters using lactose permease (LacY) from *E. coli* as a model system. We have been using

an inactive mutant enzyme for earlier studies but recently succeeded to reveal the structure of the wild type enzyme. We are going to update the current status of the study using this and earlier structures of LacY in comparison with other transporter structures. Eukaryotic membrane proteins are often difficult to produce in large quantities, which is a significant obstacle for further structural and biochemical investigation. Recently, we have reported a fluorescent-based highthroughput approach for rapidly screening membrane proteins that can be overproduced to levels of >1 mg/l in Saccharomyces cerevisiae. We find that 70% of the well-expressed membrane proteins tested in this system are stable, targeted to the correct organelle, and monodispersed. In the workshop, we will present the results of the application of this method to the production of various mammalian transporters, which are successfully purified in large quantity. We could also show that the system can, in fact, produce active mammalian transporters. We will discuss the application of this system to functional and structural studies of mammalian transporters.

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### S3/3 Crystal structure based study of NhaA, a $Na^+/H^+$ antiporter

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The Na<sup>+</sup>/H<sup>+</sup> antiporter NhaA is indispensable for pH and Na<sup>+</sup> homeostasis in Escherichia coli and many other bacteria. It has unique properties; in addition to being a transporter it has a capacity to sense the environmental signals, Na+ and H+ and to transduce the signals into a change in activity so as to maintain homeostasis. Whereas, the response to Na<sup>+</sup> occurs at the transcription level, the response to H<sup>+</sup> is conducted by the protein itself. Similar to many prokaryotic and eukaryotic antiporters NhaA is tightly regulated by pH. The crystal structure of NhaA has provided insights into the mechanism of NhaA and its unique regulation by pH. Being a novel fold, it has also shed light on the architecture of membrane transport proteins and provided a basis to intelligently design experiments both in-silico and in the molecule to study the mechanism of an antiporter and its regulation. The aim of this lecture is to describe this enlightening encounter between the crystal structure and the molecular membrane biology of NhaA.

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## S3/4 Substrate recognition and transport mechanism of mitochondrial carriers

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Mitochondrial carriers transport nucleotides, co-factors and metabolic intermediates across the mitochondrial inner membrane. The significant sequence conservation in the mitochondrial carrier family suggests that the specific recognition of substrates is coupled to a common mechanism of transport. By using two different approaches a common substrate binding site was identified consisting of residues that are highly conserved and

essential for function. The first approach uses comparative structural models and chemical and distance constraints to identify a substrate binding site capable of discriminating different substrates. The second exploits the principle that mitochondrial carriers have a high degree of three-fold pseudosymmetry in contrast to the transported substrates that are asymmetric in structure. Therefore, the substrate binding site must contain asymmetric and conserved residues to couple the binding of the asymmetric substrate to a symmetric transport mechanism. A symmetry score based on sequence comparisons was devised to asses the degree of symmetry and conservation in the carriers. Conserved asymmetry residues are found predominantly in the cavity at the midpoint of the membrane in agreement with the first approach. The common substrate binding site explains substrate selectivity, ion coupling and the effects of the membrane potential on transport. In addition, the symmetry analysis has identified residues that are important for the transport mechanism of mitochondrial carriers.

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# S3/5 Control and effect of UCP1 activity in brown-fat cells and mitochondria, and in mice and men

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Regulation of UCP1 activity has both biochemical and applied interest. However, the results are dependent upon the level of biological integration used to examine UCP1 activity.

Thus, in the simplest systems - black-lipid membranes and reconstituted vesicles - fatty-acids activate dependent upon e.g. degree of unsaturation, they have to be flipflopable and ROS products seem to affect activity. When UCP1 is studied in brownadipose-tissue mitochondria, the results are different: fatty-acids reactivate GDP-inhibited UCP1 but in an unsaturation-independent manner, they need not be flipflopable and ROS products do not affect. When hyperactivation of uninhibited UCP1 is studied, similar results are obtained. When UCP1 is studied within brown-fat cells. it can be (re)activated indirectly by norepinephrine or directly by fatty-acids. The fatty-acids need not be metabolizable, at least not beyond the acyl-CoA level. When UCP1 activity is studied within intact animals (mice), the outcome depends on temperature conditions: at normal animal house conditions, nearly no effects are seen; in the cold, nonshivering thermogenesis cannot be induced, and at thermoneutrality, UCP1 may control body weight, i.e. its absence leads to obesity. Although UCP1 was thought until recently to be absent in adult humans, its presence is now evident, and through FDG-PET its activity can be studied and correlated with physiological conditions.

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#### S3/6 A novel potassium channels in mitochondria

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